AccuDiag™
AFP (Alpha Fetoprotein)
ELISA Kit

REF 5101-16

AFP (Alpha Fetoprotein) ELISA	
Method	Enzyme Linked Immunosorbent Assay
Principle	Sandwich Complex
Detection Range	0-300ng/mL
Sample	20 µL Serum
Specificity	98.5 %
Sensitivity	2.0ng/mL
Incubation Time	80 minutes
Shelf Life	12 Months from the manufacturing date

PRODUCT FEATURES

Very easy to use with little training

Highly specific and consistent Assay

Provides accurate results quickly

Reading of results both visually and as absorbance data

INTENDED USE

AFP Enzyme Immunoassay test kit is intended for the quantitative determination of AFP concentration in human serum.

SIGNIFICANCE AND SUMMARY

Alpha-fetoprotein (AFP) is a glycoprotein with a molecular weigh of approximately 70,000 daltons. AFP is normally produced during fetal and neonatal development by the liver, yolksac, and in small concentrations by the gastrointestinal tract. After birth, serum AFP concentrations decrease rapidly, and by the second year of life and thereafter only trace amounts are normally detected in serum.

Elevation of serum AFP to abnormally high values occurs in several malignant diseases, most notably nonseminomatous testicular cancer and primary hepatocellular carcinoma. In the case of nonseminomatous testicular cancer, a direct relationship has been observed between the incidence of elevated AFP levels and the stage of disease. Elevated AFP levels have also been observed in patients diagnosed with seminoma with nonseminomatous elements, but not in patients with pure seminoma.

In addition, elevated serum AFP concentrations have been measured in patients with other noncancerous diseases, including ataxia telangiectasia, hereditary tyrosinemia, neonatal hyperbilirubinemia, acute viral hepatitis, chronic active hepatitis, and cirrhosis. Elevated serum AFP concentrations are also observed in pregnant women. Therefore, AFP measurements are not recommended for use as a screening procedure to detect the presence of cancer in the general population.

ASSAY PRINCIPLE

The AFP Quantitative Test Kit is based on a solid phase enzyme-linked immunosorbent assay. The assay system utilizes one anti-AFP antibody for solid phase (microtiter wells) immobilization and another mouse monoclonal anti-AFP antibody in the antibody-enzyme (horseradish peroxidase) conjugate solution. The test specimen (serum) is added to the AFP antibody coated microtiter wells and incubated with the Zero Buffer. If human AFP is present in the specimen, it will combine with the antibody on the well. The well is then washed to remove any residual test specimen, and AFP antibody labeled with horseradish peroxidase (conjugate) are added. The conjugate will bind immunologically to the AFP on the well, resulting in the AFP molecules being sanwiched between the solid phase and enzyme-linked antibodies. After incubation at room temperature, the wells are washed with water to remove unbound labeled antibodies. A solution of TMB is added and incubated for 20 minutes, resulting in the development of a blue color. The color development is stopped with the addition of 2N HCl, and the color is changed to yellow and measured spectrophotometrically at 450 nm. The concentration of AFP is directly proportional to the color intensity of the test sample.

SPECIMEN COLLECTION AND PREPARATION

Serum should be prepared from a whole blood specimen obtained by acceptable medical techniques. This kit is for use with serum samples without additives only.

MATERIALS AND COMPONENTS

Materials provided with the test kit

1. Antibody-coated microtiter plate with 96 wells.
2. Zero buffer, 12 ml.
3. Reference standard set, contains 0, 5, 20, 50, 150, and 300 ng/ml (WHO, 72/225) AFP, in liquid from (ready to use) or lyophilized from.
4. Enzyme Conjugate Reagent, 18 ml.
5. TMB Substrate, 12 ml.
6. Stop Solution, 12 ml.
7. Wash Buffer Concentrate (50X), 15ml
8. Controls set (optional)

Materials required but not provided

1. Precision pipettes: 5~40 µl, 50~200 µl and 1.0 ml.
2. Disposable pipette tips.
3. Distilled water.
4. Vortex mixer or equivalent.
5. Absorbent paper or paper towel.

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6. Graph paper.
7. Microtiter plate reader.

REAGENT PREPARATION

1. All reagent should be brought to room temperature (18-22°C) before us.
2. If reference standards are lyophilized, reconstitute each standard with 0.5 ml distilled water. Allow the reconstituted material to stand for at least 20 minutes. Reconstituted standards should be sealed and stored at 2-8°C.
3. Dilute 1 volume of Wash Buffer Concentrate (50x) with 49 volumes of distilled water. For example, dilute 15 ml of Wash Buffer Concentrate (50x) into 735 ml of distilled water to prepare 750 ml of washing buffer (1x). Mix well before use.

ASSAY PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense **20**μl of standard, specimens, and controls into appropriate wells.
3. Dispense **100**μl of zero buffer into each well.
4. Thoroughly mix for 10 seconds. It is very important to have complete mixing in this setup.
5. Incubate at room temperature (18-22°C) for 30 minutes.
6. Remove the incubation mixture by flicking plate content into a waste container.
7. Rinse and flick the microtiter wells 5 times with washing buffer (1X).
8. Strike the wells sharply onto absorbent paper or paper towels to remove all residual water droplets.
9. Dispense **150**μl of Enzyme Conjugate Reagent into each well. Gently mix for 5 seconds.
10. Incubate at room temperature for 30 minutes.
11. Remove the incubation mixture by flicking plate contents into a waste container.
12. Rinse and flick the microtiter wells 5 times with washing buffer (1X).
13. Strike the wells sharply onto absorbent paper to remove residual water droplets.
14. Dispense **100**μl TMB substrate into each well. Gently mix for 5 seconds.
15. Incubate at room temperature for 20 minutes.
16. Stop the reaction by adding **100**μl of stop solution to each well.
17. Gently mix for 30 seconds to make sure that the blue color changes to yellow color completely.
18. Read optical density at 450nm with a microtiter reader within 15 minutes.

Important Note:
The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

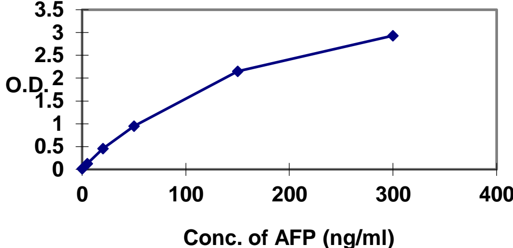
RESULTS

Calculate the mean absorbance value (A_{450}) for each set of reference standards, specimens, controls and patient samples. Constructed a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in ng/ml on graph paper, with absorbance values on the vertical or Y-axis and concentrations on the horizontal or X-axis. Use the mean absorbance values for each specimen to determine the corresponding concentration of AFP in ng/ml from the standard curve.

Example of standard curve

Results of typical standard run with optical density reading at 450nm shown in the Y-axis against AFP concentrations shown in the X-axis. This standard curve is for the purpose of illustration only, and should not be used to calculate unknowns. Each user should obtain his or her own data and standard curve.

AFP (ng/ml)	Absorbance (450nm)
0	0.012
5	0.127
20	0.455
50	0.952
150	2.150
300	2.932



EXPECTED VALUES AND SENSITIVITY

In high-risk patients, AFP values between 100 and 350 ng/ml suggest a diagnosis of hepatocellular carcinoma, and levels over 350 ng/ml usually indicate the disease. Approximately 97% of the healthy subjects have AFP levels less than 8.5 ng/ml. It is recommended that each laboratory establish its own normal range. The minimum detectable concentration of AFP by this assay is estimated to be 2.0 ng/ml.

PERFORMANCE CHARACTERISTICS

I. Accuracy

Comparison between Our Kits and Commercial Available Kits provides the following data

N = 79
Correlation Coefficient = 0.985
Slope = 1.038
Intercept = 0.729
Mean (Our) = 55.12
Mean (Abbott) = 52.24

II. Precision

1. Intra-Assay:

Concentrations	Replicates	Mean	S.D.	% CV
Level I	24	31.04	1.45	4.66
Level II	24	126.8	5.20	4.10
Level III	24	270.8	12.93	4.78

2. Inter-Assay:

Concentrations	Replicates	Mean	S.D.	% CV
Level I	24	30.58	1.88	6.1
Level II	24	125.1	7.08	5.7
Level III	24	268.3	14.06	5.2

III. Linearity

Two patient sera were serially diluted with 0 ng/mL standard in a linearity study. The average recovery was 102.2 %.

Dilution	Expected	Observed	% Recov.
undiluted	271.20	271.20	
2x	135.60	137.12	101.1
4x	67.80	69.45	102.4
8x	33.90	35.16	103.7
16x	16.95	17.20	101.5
32x	8.48	9.31	109.9
		Average Recovery: 102.2 %	

IV. Recovery

Various patient serum samples of known AFP levels were mixed and assayed in duplicate. The average recovery was 101.1 %.

Expected Concentration	Observed Concentration	% Recovery
92.70	94.20	101.6
77.63	79.31	102.2
23.24	23.10	99.4
156.18	159.23	102.0
206.06	210.10	102.0
148.94	150.52	101.1
149.73	148.97	99.50
	Average Recovery: 101.1 %	

V. Sensitivity

The minimum detectable concentration of this assay is estimated to be 2.0 ng/mL.

VI. Cross-reactivity

The following human materials were tested for crossreactivity of the assay:

Antigens	Concentration	Equivalent AFP	% Cross-reactivity
hCG	400 IU/mL	0.00	0.00
PAP	1,000 ng/mL	0.00	0.00
PSA	1,000 ng/mL	0.00	0.00
CEA	1,000 ng/mL	0.00	0.00

VII. Hook Effect

No hook effect was observed in this assay.

LIMITATIONS OF THE PROCEDURE

There are some limitations of the assay:

1. As with all diagnostic tests, a definite clinical diagnosis should not be based on the results of a single test, but should only be made by the physician after all clinical and laboratory findings have been evaluated.
2. Studies have implicated possible interference in immunoassay results in some patients with known rheumatoid factor and antinuclear antibodies. Serum samples from patients who have received infusions containing mouse monoclonal antibodies for diagnostic or therapeutic purposes, may contain antibody to mouse protein (HAMA). Although we have added some agents to avoid the interferences, we cannot guarantee it will eliminate all the effects of that.

STORAGE

1. Unopened test kits should be stored at 2-8°C upon receipt.
2. The microtiter plate should be kept in a sealed bag with desiccants to minimize exposure to damp air.
3. Opened test kits will remain stable until the expiring date shown, provided it is stored as prescribed above.
4. A microtiter plate reader with a bandwidth of 10nm or less and an optical density range of 0-2 OD or greater at 450nm wavelength is acceptable for use in absorbance measurement.

REFERENCES

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MANUFACTURER AND BRAND DETAILS

ISO 13485:2016

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Date Adopted	2022-09
Brand Name	AccuDiag™
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EC REP	CEpartner4U, Esdoornlaan 13, 3951 DB Maarn, The Netherlands. www.cepartner4u.eu
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